

## Modulating and Understanding Retention of Proteins on Chromatographic Support by Changing Cation-Exchanging Ligand

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### 1. Introduction

Ion-exchange chromatography is a key chromatographic technique for the separation and purification of large biomolecules; however, its performance strongly depends on the nature of the ligand and the underlying support matrix. Our research investigates the effects of sulfonate (CIM<sup>+</sup> SO<sub>3</sub>) and multimodal cation-exchange sulphate (CIM SO<sub>4</sub>) ligands on the chromatographic separation of two basic proteins: equine cytochrome c (Cyt) and chicken egg-white lysozyme (Lys).

When performing protein separation under an ascending salt gradient, the elution of both Cyt and Lys were dramatically shifted to higher NaCl on sulfate ligand. The effect was more pronounced for Lys despite their comparable molecular sizes and isoelectric points (pI). Since Lys is reported to be slightly more hydrophobic than Cyt, we hypothesize that CIM SO<sub>4</sub>, due to reported higher hydrophobicity of SO<sub>4</sub> groups, interacts with Lys stronger through additionally expressed "hydrophobic" interactions [1, 2].

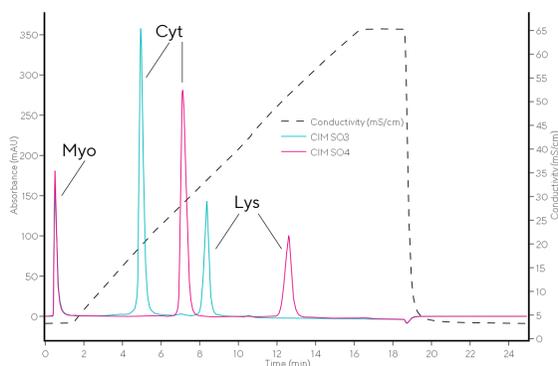


Figure 1: Separation of myoglobin (Myo), Cyt and Lys on CIM SO<sub>3</sub> and on CIM SO<sub>4</sub> columns in NaCl gradient.

Buffer A: 20mM phosphate, pH 7.5

Buffer B: 20mM phosphate, 0.7 M NaCl, pH=7.5

Detection: UV at 280 nm and conductivity

Method: Linear ascending NaCl gradient from 100% buffer A to 100% buffer B over 30 CV

### 2. Experimental Approach

The goal of this study was to describe the chromatographic differences by performing and interpreting thermodynamic analyses for Lys and Cyt adsorption at different binding conditions. Two complementary methods were used: (i) chromatographic analysis using the Van't Hoff equation, and (ii) isothermal titration calorimetry (ITC).

(i) Van't Hoff thermodynamic analysis of retention data

Protein	Cytochrome	Lysozyme
Column	CIM SO <sub>3</sub> and CIM SO <sub>4</sub>	
Mode	isocratic	
Temp.	25, 30, 35, 40 °C	
Buffer	20 mM phosphate, 0.15 M NaCl, pH 7.5	20 mM phosphate, 0.29 M NaCl, pH 7.5
Sample	0.60 mg/mL myoglobin, 0.75 mg/mL Cyt in the Buffer	0.70 mg/mL Lys in the Buffer

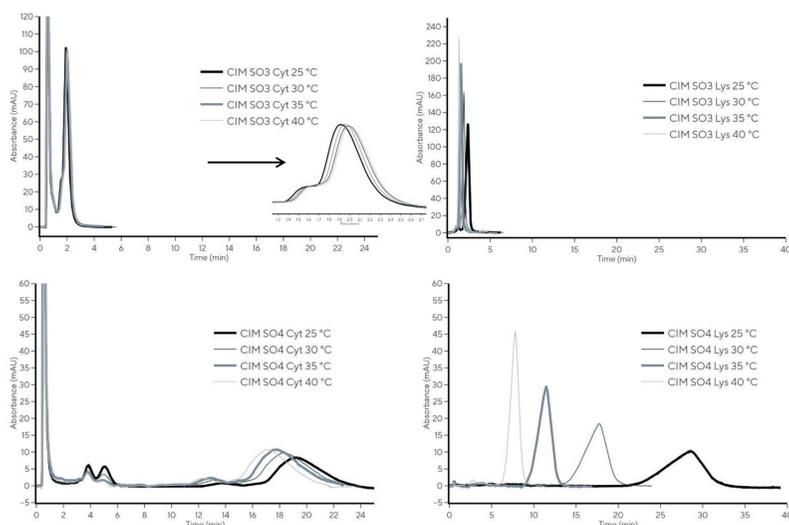


Figure 2: Chromatograms obtained at 25, 30, 35 and 40 °C injecting Myo (nonbinding tracer) with Cyt or Lys onto a CIM SO<sub>3</sub> and CIM SO<sub>4</sub> under isocratic chromatographic conditions.

The Van't Hoff equation enables the calculation of thermodynamic binding parameters directly from the retention time of the protein:

$$\ln k' = a + \frac{b}{T} + \frac{c}{T^2} + \ln \varphi$$

$$\Delta H^\circ = -R \left( b + \frac{2c}{T} \right)$$

$$\Delta S^\circ = R \left( a - \frac{c}{T^2} \right)$$

$$\Delta C_p^\circ = \frac{2Rc}{T^2}$$

Where  $k'$  is the retention factor,  $T$  the absolute temperature and  $\varphi$  the phase ratio,  $a$ ,  $b$  and  $c$  represent empirical coefficients, which were used to calculate the thermodynamic parameters [3].

(ii) Isothermal titration calorimetry (ITC)

ITC experiments were performed using a 1 mL sample cell containing a slurry of the ground monolithic stationary phase, while the 250  $\mu$ L syringe was filled with protein solution (5 mg/mL Lys in 20 mM phosphate, 0.10 M NaCl, pH 7.5). Titrations were carried out at 25 °C. For each measurement, 25 injections of 10  $\mu$ L were performed under continuous stirring.

The power (heat rate) required to compensate for the heat released or absorbed upon each injection of protein into the ligand solution was recorded and integrated (raw signal shown in Figure 3). The resulting heats (Figure 4) were analyzed using Wiseman fitting to determine the thermodynamic binding parameters.

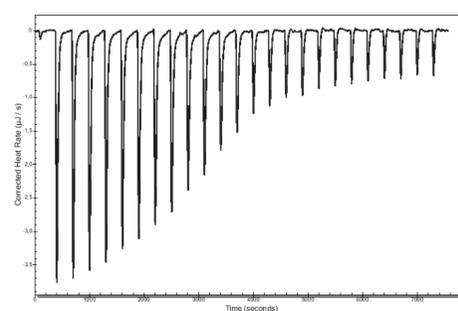


Figure 3: Raw ITC signal obtained for titration of Lys sample solution into 5% CIM SO<sub>3</sub> slurry at 25 °C

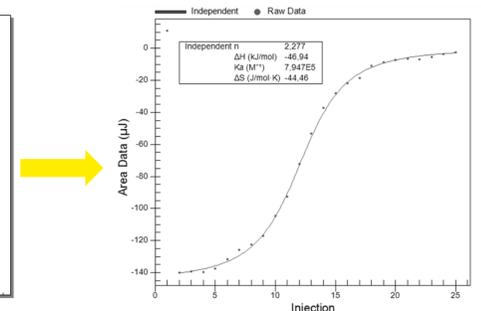
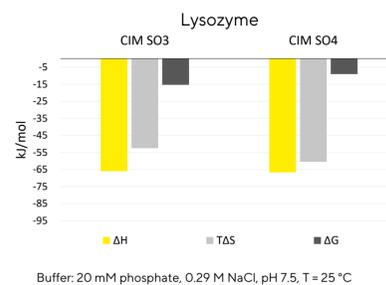


Figure 4: Titration curve for CIM SO<sub>3</sub> slurry

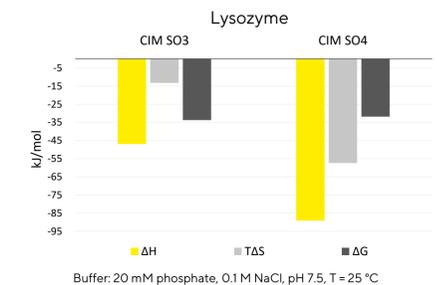
### 3. Results

Van't Hoff



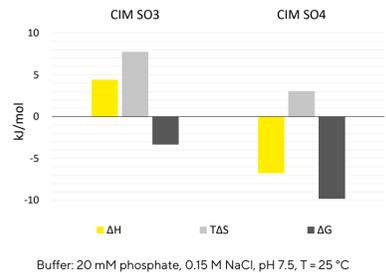
Buffer: 20 mM phosphate, 0.29 M NaCl, pH 7.5, T = 25 °C

ITC



Buffer: 20 mM phosphate, 0.1 M NaCl, pH 7.5, T = 25 °C

Cytochrome c



Buffer: 20 mM phosphate, 0.15 M NaCl, pH 7.5, T = 25 °C

Lys	Cyt
CIM SO <sub>3</sub> and SO <sub>4</sub> :	CIM SO <sub>3</sub> :
<ul style="list-style-type: none"> <li>enthalpy-driven process</li> <li>Strongly exothermic process, dominated by electrostatic interactions</li> <li>negative <math>\Delta S</math> values <math>\rightarrow</math> hydrogen bonds formation and a decrease in conformational freedom upon adsorption.</li> </ul>	<ul style="list-style-type: none"> <li>entropy-favorable process</li> <li>endothermic process + high positive <math>\Delta S \rightarrow</math> disruption of hydrogen bonds and release of bound water molecules</li> </ul>
	CIM SO <sub>4</sub> :
	<ul style="list-style-type: none"> <li>exothermic process + positive <math>\Delta S \rightarrow</math> ionic interaction + dehydration</li> </ul>

Van't Hoff vs ITC analysis of Lys adsorption

- Both Van't Hoff and ITC analyses confirmed enthalpy-driven lysozyme binding to both cation-exchange monoliths, indicating the domination of attractive electrostatic and hydrogen-bonding forces.
- The Van't Hoff analysis revealed only minor differences between the two ligands, with slightly stronger enthalpic contribution observed for the multimodal CIM SO<sub>4</sub> ligand, while ITC clearly discriminates between the two exchangers. Possible reason 1: Due to its indirect nature and the assumption of ideal retention behavior, the Van't Hoff approach provides a less sensitive differentiation between binding modes. Possible reason 2: The two methods were performed at different salt concentrations (ITC at 0.1 M NaCl, Van't Hoff at 0.29 M), affecting the mechanism of adsorption.
- ITC data reveal that the stronger binding on the multimodal SO<sub>4</sub><sup>-</sup> ligand originates from intense ionic interactions and specific hydrogen bonding between the arginine residues of lysozyme and the negatively charged sulfate groups. The difference in total heat release between SO<sub>3</sub><sup>-</sup> and SO<sub>4</sub><sup>-</sup> ligands reflects the additional stabilizing interactions provided by the multimodal ligand.

### 4. Conclusion

- Under identical isocratic conditions, Lys showed a 12-times higher retention time using CIM SO<sub>4</sub> compared to CIM SO<sub>3</sub>.
- Van't Hoff thermodynamic analysis revealed no major differences between the exchangers for Lys, whereas Cyt exhibited a distinct adsorption mechanism depending on the ligand type.
- ITC clearly differentiated the ion exchangers: CIM SO<sub>3</sub> demonstrated a more entropy-driven adsorption, while CIM SO<sub>4</sub> showed a higher negative  $T\Delta S$ , indicating a more enthalpy-dominated interaction. Based on these unexpected observations, we propose that protein adsorption is influenced not only by the ligand chemistry, but that the backbone of the matrix (polymethacrylate polymer) itself plays an equally important role in determining the binding mechanism.

### 5. References

- [1] J. C. Simoes-Cardoso et al. Langmuir 2020, 36, 3336-3345. <https://doi.org/10.1021/acs.langmuir.0c00197>
- [2] Begič, M. Et al. Biotechnol. J. 2021, 16, e2100100. <https://doi.org/10.1002/biot.202100100>
- [3] F.S. Marquesa et al. Colloids and Surfaces B: Biointerfaces 2014, 122, 801-807. <https://doi.org/10.1016/j.colsurfb.2014.08.024>