

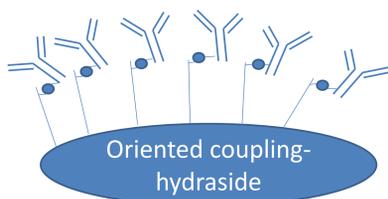
Isolation of RAE-1 from murine cells based on immunoaffinity monolithic chromatography

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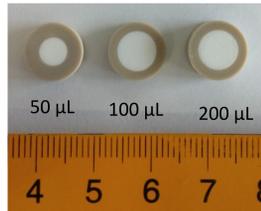
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INTRODUCTION

Immunoaffinity columns using antibodies as ligands against mammalian proteins could be used for different applications in protein expression control and, if a standard available, for direct protein quantification in complex sample solutions. Additionally, these columns are ideal for polishing step of recombinant proteins, such as mammalian receptor Fc fusion proteins. Most importantly, such columns could extract a significant amount of a single membrane protein from native source, suitable for downstream analyses, such as mass spec analysis of their glycans. Immunoaffinity chromatographic monoliths against RAE-1 GPI anchored glycoprotein were developed (CIMmic α RAE-1 column) as a part of Glycomet project with the main goal to analyze the antigen glycoprofile.



CIMmic chromatographic support (white circles):



CIMmic housing:

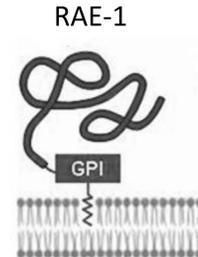


Figure 2: RAE-1 is a GPI anchored protein. To solubilize hydrophobic proteins, detergent is needed before and during separation.

Figure 1: Antibodies immobilized on monoliths in oriented direction. On hydrazide preactivated support (HIDA), monoclonal antibodies (mAbs) are immobilized through oxidized glycans. CIMmic[®] monolithic disks with column volume of 50 μ L were used as a chromatographic support for immunoaffinity chromatography

RESULTS

Antibodies

Figure 3: α RAE-1 mAb precipitates RAE-1 from cell lysates of corresponding transfectants. RAE-1 is heavily glycosylated and thus forms smear in WB.

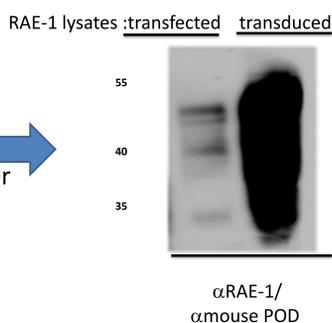
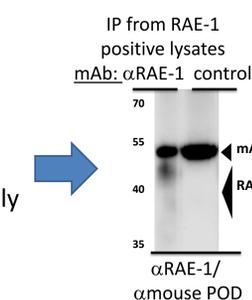


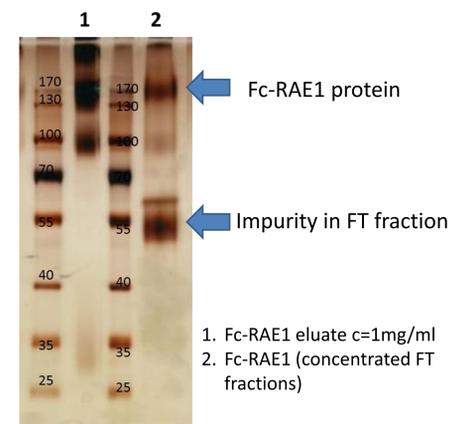
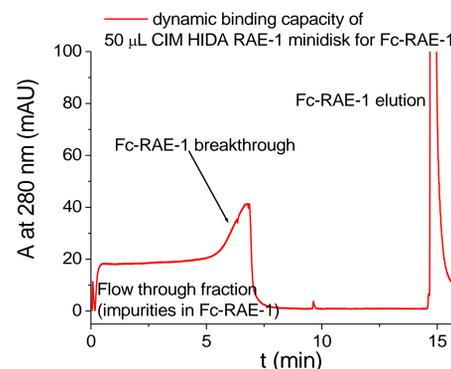
Figure 4: The amount of proteins upon transduction (using a cytomegaloviral vector) is significantly higher than in transfected cells (RAE-1)

CIMmic α RAE-1 column

α RAE-1 mAb immobilisation: Immobilisation procedure was a platform one, developed before for other IgGs. Approximately 500 μ g of IgG is needed for a 50 μ L minidisk.

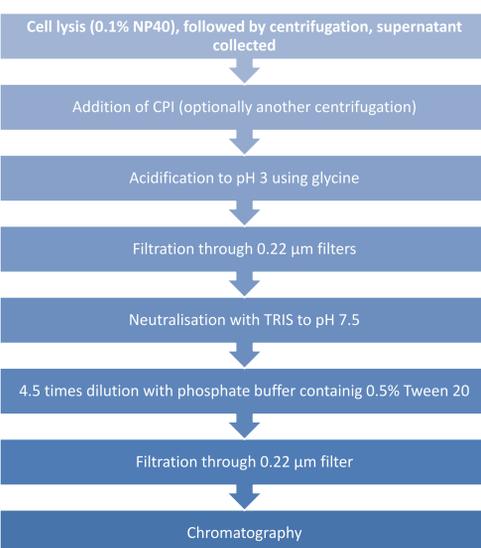
Figure 5: Dynamic binding capacity for Fc-RAE-1 on CIMmic α RAE-1 column was determined to be cca 5 mg Fc-RAE-1 per mL of chromatographic support.

A strong signal in FT fraction was proven to be an impurity (see SDS-PAGE on the right) after standard purification protocol for recombinant Fc fusion proteins. This opens up new perspectives to obtain high-quality protein samples needed for certain functional and immunological assays.

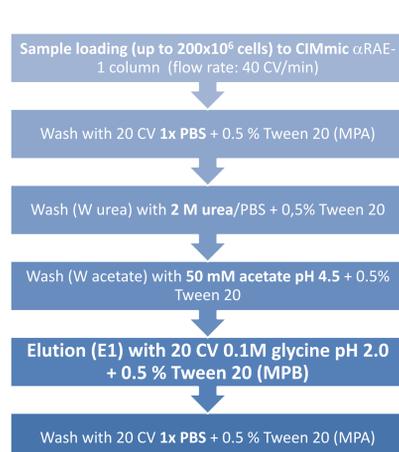


RAE-1 isolation from cells

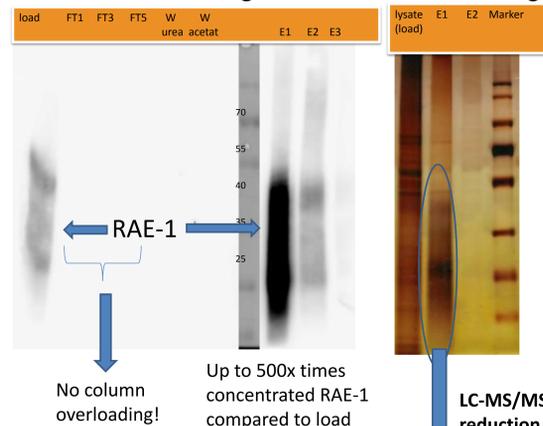
Sample preparation:



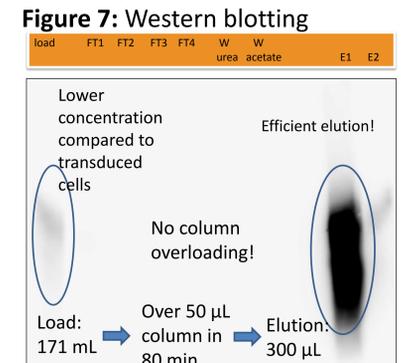
Chromatography:



RAE-1 isolation from transduced NIH cells; Figure 6: Western blotting



RAE-1 isolation from 160 million transfected NIH cells; Figure 7: Western blotting



Accession	Description	Score	Coverage	# Proteins	# Unique peptides	MW (kDa)
O08604	Retinoic acid early-inducible protein 1-gamma	6162	32.02	1	5	28.5

CONCLUSIONS

- Transduced cells are ideal for massive production of mammalian membrane glycoproteins.
- Hydrazide coupling chemistry was proven as optimal for preparation of immunoaffinity α RAE-1 monolithic chromatographic support.
- Microgram amounts of low-abundant native RAE-1 protein were purified from cell lysates.
- The protein sample preparation allows N-glycosylation analysis of purified RAE-1.
- The N-linked glycosylation comparison of transfected and transduced proteins might elucidate the mechanism exploited by viruses to evade immune response.